## New Disease Reports First finding of Phytophthora foliorum in the United Kingdom

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In January 2016, Science and Advice for Scottish Agriculture received shoots of Rhododendron ponticum submitted by a Forestry Commission Scotland Tree Health Officer. The samples had been collected during a survey for P. ramorum from an extensive area of wild rhododendron growing along a road in the northwest of Scotland (Fig. 1).

Affected shoots were wilted with red-purple discoloration of the leaves as well as dieback of the branch tips. Segments of leaves and stems bordering the discolorations and dieback were plated onto V8 agar containing antibiotics selective for Phytophthora spp. (PARPNH; Jung et al., 1996). The agar plates were incubated at 18°C and after five days a Phytophthora sp. had grown from some of the segments. The resulting culture was homothallic with an abundance of plerotic, thick-walled oogonia and predominantly paragynous antheridia (Fig. 2). No sporangia were observed. The growth rate on V8 agar was 51-56 mm after seven days at 20°C with little aerial mycelium (Fig. 3).

Sequencing the ITS region of the ribosomal RNA gene (GenBank Accession No. KX364273) after amplification with primers ITS4 and ITS6 (White et al., 1990; Cooke & Duncan, 1997) revealed a 100% match with four isolates of P. foliorum (e.g. KJ755120 and EF120469). A subsequent sample from the same plant was sent to the Tree Health Diagnostic and Advisory Service of Forest Research, Alice Holt where the finding was confirmed.

To complete Koch's postulates the abaxial sides of detached R. ponticum 'Variegatum' leaves (n=12) were inoculated with agar plugs (4 mm diameter) from a two-week-old P. foliorum culture in a 50 µl droplet of sterile distilled water that was replenished every two-three days. One half of the leaves was wounded by a pin prick before inoculation the other half was left non-wounded. Leaves were incubated at 20°C under natural light. After seven days, lesions (1-1.5 cm diameter) developed around the wounds from which the pathogen was readily re-isolated; without wounding, no lesions formed. For comparison, the same host was inoculated with other Phytophthora spp. at the same time. P. foliorum appeared to be less aggressive than P. kernoviae and P. ramorum, but more aggressive than P. gonapodyides and P. syringae (Fig. 4).

Phytophthora foliorum was first described by Donahoo et al. (2006) attacking the foliage of evergreen hybrid azaleas (Rhododendron sp.) in nurseries in California and Tennessee, USA. No other hosts have been reported to date. The only other report of this pathogen is from two-yearold azalea plants in two container stands from an ornamental nursery in Spain (Jung et al., 2016). The finding in Scotland is the first detection in the UK and the first finding in the wider environment in the world. A follow-up survey in the surrounding area found a second wild R. ponticum with P. foliorum which was also infected by P. ramorum. Other Phytophthora species present on R. ponticum in the surrounding area were P. gonapodyides and P. syringae. All infected plants have now been removed as part of a rhododendron clearance programme.

## References

Cooke DEL, Duncan JM, 1997. Phylogenetic analysis of Phytophthora species based on ITS1 and ITS2 sequences of the ribosomal RNA gene repeat. Mycological Research 101, 667-677. http://dx.doi.org/10.1017/S0953756296003218

Donahoo R, Blomquist CL, Thomas SL, Moulton JK, Cooke DEL, Lamour, KH, 2006. Phytophthora foliorum sp. nov., a new species causing leaf blight of azalea. Mycological Research 110, 1309-1322. http://dx.doi.org/10.1016/j.mycres.2006.07.017

Jung T, Blaschke H, Neumann P, 1996. Isolation, identification and pathogenicity of Phytophthora species from declining oak stands. European Journal of Forest Pathology 26, 253-272.

http://dx.doi.org/10.1111/j.1439-0329.1996.tb00846.x

Jung T et al., 2016. Widespread Phytophthora infestations in European nurseries put forests, semi-natural and horticultural ecosystems at high risk of Phytophthora diseases. Forest Pathology 46, 134-163. http://dx.doi.org/10.1111/efp.12239

White TJ, Burns T, Lee S, Taylor J, 1990. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis MA, Gelfand DH, Sninsky JJ, White TJ eds. PCR Protocols - A Guide to Methods and Applications. London, UK, Academic Press, 315-322.

Figure 1





Figure 2





Figure 3

## Figure 4

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