# New Disease Reports

## First report of *Colletotrichum gloeosporioides* species complex causing anthracnose on leaves of cutnut, *Barringtonia edulis*, in Papua New Guinea

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Received: 10 Jan 2017. Published: 07 Feb 2017. Keywords: foliar fungal plant disease

Cutnut, *Barringtonia edulis*, is one of six indigenous edible nut tree species found in Papua New Guinea (PNG), Solomon Islands and Vanuatu, where it has market potential both for domestic consumption and as an export commodity (Wah, 1996). Here we report an anthracnose-like foliar disease observed in January 2015 on 90% (19 out of 21) cutnut trees planted for landscaping at the PNG University of Technology, Lae. Foliar symptoms gradually progressed from an angular, asymmetrical yellow discoloration to an ulcer-like necrosis causing death to the whole leaf (Fig. 1).

Portions of necrotic leaf tissues (1 cm<sup>2</sup>) were excised and surface-sterilised with 5g/l NaOCl for five minutes, rinsed three times with sterilised water and plated onto fresh potato dextrose agar (PDA) then incubated at 25°C in the dark. A pure culture was obtained using a modified monoconidial technique by slightly agitating seven-day-old cultures over fresh PDA to release the spores under laminar airflow conditions; the resulting sporeinoculated cultures were then incubated in the dark at 25°C. After 24-48 hours, germinating spores were identified under a dissecting light microscope and transferred onto PDA using a sterile glass needle. Cultural and morphological descriptions were done after 14 days incubation at 25°C in the dark. This methodology is as described by Phoulivong et al. (2012). Colony pigmentations observed on PDA ranged from grey to white with slightly raised aerial mycelium to a dense cottony mycelium. Cultures exhibited salmon to bright orange acervuli arranged in concentric rings (Fig. 2), with aging the acervuli become evenly distributed towards the edges of the culture (Fig. 3). The conidial shape was straight and cylindrical with the average conidia length and width ranging from 12-15.7 and 4-5.3 µm respectively, fitting published descriptions (Photita et al., 2005; Weir et al., 2012) (Fig. 4). DNA-based PCR amplification using the forward species-specific primer, 5'-GGGGAAGCCTCTCGCGG-3' (Mills et al., 1992), and universal ITS4 (reverse) primer targeting the ribosomal DNA of Colletotrichum gloeosporioides produced a 450 bp amplicon. A BLAST search found the amplicon sequence (GenBank Accession No. KT601169) to have 99% identity to several C. gloeosporioides accessions (Table 1).

Koch's postulates were fulfilled using 10 detached fifth-largest, fullyopened *B. edulis* leaves from healthy plants raised in a greenhouse which were inoculated with 0.5 ml of a  $10^5$  spores/ml spore suspension and incubated at 25°C in the dark. The fungus was re-isolated from anthracnoselike lesions formed on these leaves, and the identity as *C. gloeosporioides* species complex was confirmed by PCR using the same primers. Anthracnose is mainly caused by the *C. gloeosporioides* species complex (Weir *et al.*, 2012). It is reported to affect a wide range of commercially important crops (e.g. Than *et al.*, 2008). To our best knowledge, this is the first report of the *C. gloeosporioides* species complex causing foliar anthracnose on cutnut, *B. edulis*, in PNG and in the world. As such, it is imperative to identify the causal pathogen(s) of the complex to assess the potential for yield loss in commercial nut production, and to evaluate the implications for the management of the disease in other crop species.

#### Acknowledgements

This work was financially supported by the UNITECH Research Committee, Papua New Guinea University of Technology.

#### References

Mills PR, Sreenivasaprasad S, Brown AE, 1992. Detection and differentiation of *Colletotrichum gloeosporioides* isolates using PCR. *FEMS Microbiology Letters* **98**, 137-143.

http://dx.doi.org/10.1111/j.1574-6968.1992.tb05503.x

Photita W, Taylor PWJ, Ford R, Hyde KD, Lumyong S, 2005. Morphological and molecular characterization of *Collectorichum* species form herbaceous plants in Thailand. *Fungal Diversity* **18**, 117-133.

Phoulivong S, McKenzie EHC, Hyde KD, 2012. Cross infection of *Colletotrichum* species; a case study with tropical fruits. *Current Research in Environmental and Applied Mycology* **2**, 99-111. http://dx.doi.org/10.5943/cream/2/2/2

Than PP, Jeewon R, Hyde KD, Pongsupasamit S, Mongkolporn O, Taylor, PWJ, 2008. Characterization and pathogenicity of *Colletotrichum* species associated with anthracnose on chilli (*Capsicum* spp.) in Thailand. *Plant Pathology* **57**, 562-572.

### http://dx.doi.org/10.1111/j.1365-3059.2007.01782.x

Wah LC, 1996. Marketing indigenous nuts in Vanuatu - a private enterprise perspective. In: Stevens ML, Bourke RM, Evans BR, eds. *Proceedings of South Pacific Indigenous Nuts workshop*, ACIAR Proceedings No. 69, Port Vila, Vanuatu, 79.

Weir BS, Johnston PR, Damm U, 2012. The *Colletotrichum gloeosporioides* species complex. *Studies in Mycology* **73**, 115-180. http://dx.doi.org/10.3114/sim0011

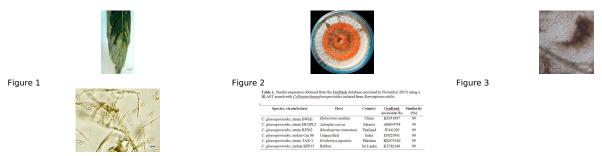


Figure 4

Figure 5

**To cite this report**: Buyoyu P, Maino MK, Okpul T, 2017. First report of *Colletotrichum gloeosporioides* species complex causing anthracnose on leaves of cutnut, *Barringtonia edulis*, in Papua New Guinea. *New Disease Reports* **35**, 7. http://dx.doi.org/10.5197/j.2044-0588.2017.035.007

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