

Natural occurrence of *Malva vein clearing virus* in malva in Iran

H. Valouzi¹, A. Golnaraghi²* and F. Rakhshandehroo¹

¹ Department of Plant Pathology, College of Agriculture and Natural Resources, Science and Research Branch, Islamic Azad University (IAU), P.O. Box 14515-775, Tehran, Iran; ² Department of Plant Protection, College of Agriculture and Natural Resources, Science and Research Branch, Islamic Azad University (IAU), P.O. Box 14515-775, Tehran, Iran

*E-mail: agolnaraghi@yahoo.com

Received: 27 Feb 2017. Published: 22 Mar 2017. Keywords: mallow

Malva plants (*Malva* spp.) are widespread in Iran and grown in different geographical regions in the country; virus-like symptoms, e.g. mosaic, are frequently observed on these plants. In the present study, eleven symptombearing malva leaf samples, showing severe and mild mosaic, vein clearing and leaf deformation, were collected from plants grown in natural ecosystems as well as in or around fields in the Markazi and Tehran provinces of Iran during the growing seasons of 2012-2014. Each leaf sample was tested for the presence of potyviruses by indirect ELISA using a "poty-group test" kit (Bioreba, Switzerland), according to the manufacturer's instructions. The ELISA results revealed a positive reaction for seven samples using the potyvirus antibodies. Mosaic, vein clearing and leaf deformation symptoms were associated with potyvirus infections.

Total RNA of three ELISA-positive samples was extracted using a LiCl protocol (Rosas-Cárdenas et al., 2011). The extracted RNAs were used as template in RT-PCR assays using universal potyvirus primers NIb2F and NIb3R (Zheng et al., 2010). The RT-PCR assays resulted in the amplification of fragments with the expected size of 350 bp, confirming the ELISA potyvirus results for all tested samples. The nucleotide sequences of the amplicons were determined using the amplification primers and deposited in GenBank (Accession Nos. KU962948, KU962952 and KU962957). BLAST analysis showed that the isolates had the highest nucleotide sequence identities (84-89%) with Malva vein clearing virus (MVCV). For further analysis, the 3' end genome sequence of isolate Ma-W2243 from Malva sp. in Delijan (Markazi province) (Fig. 1), including the partial coat protein (CP) gene sequence, was amplified using NWCIEN and Tu3T9M primers, and sequenced using the primer Nseq (Tan et al., 2005; Valouzi et al., 2017). The gap between the NIb and CP regions of the isolate was subsequently amplified using a pair of newly designed primers 5'-ATGGATTACCTGATGGTTGG-3' (F: and R: 5'-TGTCTGAACGTTGGTGCTGC-3') and sequenced to obtain the complete sequence of the CP gene. The CP sequence of the isolate was then assembled (909 nts) and deposited in GenBank (KY653927). BLAST analysis showed that the sequence had 90% nucleotide and 95% amino acid identity with sequences of MVCV. Phylogenetic analysis of the sequence and representative sequences from the genus Potyvirus also revealed a clustering of the isolate Ma-W2243 with other isolates of MVCV, confirming the identity of the virus.

To our knowledge, this study reports the natural occurrence of MVCV on plants of *Malva* spp. in Iran for the first time. MVCV is a distinct species in the genus *Potyvirus* and has a narrow host range. The virus naturally infects weeds and garden plants belonging to the family Malvaceae (Brunt *et al.*, 1996; Menzel *et al.*, 2010). Work is ongoing to investigate the presence of MVCV in other species and to study other viruses occurring in malva in the country.

Acknowledgements

We are grateful to the Science and Research Branch of IAU (Tehran) for supporting this project. This work was mainly supported by grants from the Iranian Group for the Promotion of Science, IGPS, nos. 91001003 and 92001005.

References

Brunt AA, Crabtree K, Dallwitz MJ, Gibbs AJ, Watson L, eds, 1996. Viruses of plants. Descriptions and lists from the VIDE database. Wallingford, UK: CABI.

Menzel W, Winter S, Richert-Pöggeler KR, 2010. First report of *Malva* vein clearing virus naturally occurring in hollyhock in Germany. *Plant Disease* **94**, 276. <u>http://dx.doi.org/10.1094/PDIS-94-2-0276B</u>

Rosas-Cárdenas FF, Durán-Figueroa N, Vielle-Calzada JP, Cruz-Hernandez A, Marsch-Martinez N, de Folter S, 2011. A simple and efficient method for isolating small RNAs from different plant species. *Plant Methods* **7**, 4. <u>http://dx.doi.org/10.1186/1746-4811-7-4</u>

Tan Z, Gibbs AJ, Tomitaka Y, Sánchez F, Ponz F, Ohshima K, 2005. Mutations in *Turnip mosaic virus* genomes that have adapted to *Raphanus* sativus. Journal of General Virology **86**, 501-510. http://dx.doi.org/10.1099/vir.0.80540-0

Valouzi H, Golnaraghi A, Abedini-Aminabad L, Diyanat M, 2017. Serological and molecular identification of *Turnip mosaic virus* in some wild plants in Iran. *Australasian Plant Disease Notes* **12**, 1-4. http://dx.doi.org/10.1007/s13314-016-0225-2

Zheng L, Rodoni BC, Gibbs MJ, Gibbs AJ, 2010. A novel pair of universal primers for the detection of potyviruses. *Plant Pathology* **59**, 211-220. http://dx.doi.org/10.1111/j.1365-3059.2009.02201.x



Figure 1

To cite this report: Valouzi H, Golnaraghi A, Rakhshandehroo F, 2017. Natural occurrence of *Malva vein clearing virus* in malva in Iran. *New Disease Reports* **35**, 15. <u>http://dx.doi.org/10.5197/j.2044-0588.2017.035.015</u> © 2017 The Authors *This report was published on-line at www.ndrs.org.uk where high quality versions of the figures can be found.*