



First report of *Nerine latent virus* in *Haemanthus albiflos* in the UK

W.A. Monger and R.J. Eden

Science and Advice for Scottish Agriculture (SASA), Roddinglaw Road, Edinburgh, EH12 9FJ, United Kingdom

*E-mail: wendy.monger@sasa.gsi.gov.uk

Received: 12 Nov 2018. Published: 22 Jan 2019.

Haemanthus albiflos is a flowering plant native to South Africa and Namibia. A member of the family *Amaryllidaceae*, it is an evergreen bulbous perennial with unusual flowers that give it the common name of paintbrush. Tolerance for indirect sunlight and neglect has made this a popular houseplant in countries where it would not survive the winter climate. One such house plant bought at a UK nursery in the late 1990's, was later recognised as showing mild streaking symptoms consistent with a virus infection (Fig. 1). The authors can find no record of viruses associated with *Haemanthus albiflos* but the related plant *Haemanthus multiflorus* (blood lily) recently removed from the *Haemanthus* genus and renamed *Scadoxus multiflorus*, is a host for *Nerine latent virus* (NeLV) (Chen *et al.*, 2016). From accessions on the NCBI database blood lily is also recorded as a host plant for *Cucumber mosaic virus* (CMV) and *Tomato spotted wilt virus* (TSWV).

The haemanthus plant was investigated using immuno-strips (Agdia) to detect CMV, *Impatiens necrotic spot virus*, *Iris yellow spot virus* and TSWV but no antibody reactions occurred. This was followed by total RNA extraction from leaves (Plant RNeasy kit, Qiagen) and RT-PCR with generic primers that amplify viruses from the *Carlavirus* (Nie *et al.*, 2008) and *Potexvirus* genera (van der Vlugt & Berendsen, 2002) and the *Potyviriidae* family (Gibbs & Mackenzie, 1997). The testing indicated a positive result with the carlavirus primer set Car-F2b and oligo-dT. The sequenced PCR product when compared to the NCBI database, indicated a close match with *Nerine latent virus* (NeLV), also known as Narcissus symptomless virus. To further characterise this isolate, the complete coat protein (CP) was amplified (Genbank Accession No. MK085064) using specific primers either side of the CP region (NeLV6946F 5'-AGAGTTTGTGCTCTTAGGTTA-3', NeLV8018R 5'-AAGGAGCCACTTGATTGCTT-3'). Sequence alignments, with available NeLV CP sequences (NCBI database), showed that the virus isolates shared high identity with each other, 95-98% at the nucleotide level. The highest identity of 98% was with JX524884 from narcissus (Taiwan) and HM119498 from nerine (Netherlands). The alignments revealed the haemanthus isolate had three nucleotides (one amino acid) less than other NeLV isolates and this deletion was located near the five prime end of the CP. The phylogenetic relationship of the NeLV CPs from different host plants and countries of origin are shown in Figure 2.

NeLV is reported as symptomless in the blood lily plant (Chen *et al.*, 2016) and previously in the host species of *Nerine bowdenii* and *Hippeastrum hybridum* (Brunt *et al.*, 1996) but mosaic symptoms have been reported in *Crinum* plants (Jordan *et al.*, 2018). The symptoms produced by NeLV on indicator plants have been recorded (Brunt *et al.*, 1996): local chlorotic

lesions with *Chenopodium murale* and *C. quinoa* with systemic infection only reported for *Nicotiana clelandii*; whilst *N. benthamiana* and *N. tabacum* were reported as insusceptible. To confirm this report and assess if a second virus may be present in the haemanthus, two each of the previously mentioned indicator plants were inoculated with ground leaf sap, diluted in water. Clear chlorotic symptoms were visible on the *C. quinoa* inoculated leaves from six days post inoculation (dpi), whereas symptoms on the other plants were either not present or not clear and could be mistaken for inoculation damage (Fig. 3). No systemic symptoms were seen. Subsequent RT-PCR with specific NeLV primers performed 14 dpi on the inoculated leaves found that only the *N. clelandii* gave a strong product. *N. clelandii* had shown no clear symptoms but the PCR results indicated replication and systemic movement of this virus that had not occurred in the other indicator plants. Whilst the testing was not exhaustive, the PCR, immuno- and bio-assays gave no indication that a second virus was present in the haemanthus plant and the observed symptoms are probably caused by NeLV. This report represents the first finding of NeLV in *H. albiflos*.

References

- Brunt AA, Crabtree K, Dallwitz MJ, Gibbs AJ, Watson L, eds, 1996. *Viruses of plants. Descriptions and lists from the VIDE database*. Wallingford, UK: CABI.
- Chen CC, Chang CA, Chiang FL, 2016. Serological and molecular identification of *Nerine latent virus* infecting blood lily (*Haemanthus multiflorus* Martyn). *Journal of Taiwan Agricultural Research* **65**, 194-206.
- Gibbs A, Mackenzie A, 1997. A primer pair for amplifying part of the genome of all potyvirids by RT-PCR. *Journal of Virological Methods* **63**, 9-16. [http://dx.doi.org/10.1016/S0166-0934\(96\)02103-9](http://dx.doi.org/10.1016/S0166-0934(96)02103-9)
- Jordan R, Wingert M, Loudon C, Guaragna MA, 2018. First report of *Nerine latent virus* in ornamental *Crinum* in the United States. *Plant Disease* **102**, 1469. <http://dx.doi.org/10.1094/PDIS-09-17-1512-PDN>
- Nie X, Bai Y, Molen TA, Desjardins DC, 2008. Development of universal primers for detection of potato carlaviruses by RT-PCR. *Journal of Virological Methods* **149**, 209-216. <http://dx.doi.org/10.1016/j.jviromet.2008.02.004>
- van der Vlugt RAA, Berendsen M, 2002. Development of a general potexvirus detection method. *European Journal of Plant Pathology* **108**, 367-371. <http://dx.doi.org/10.1023/A:1015644409484>



Figure 1

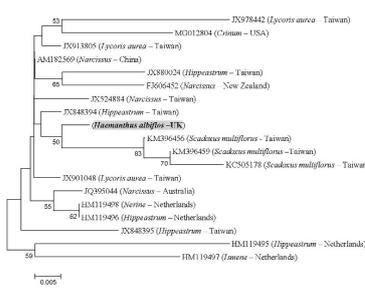


Figure 2



Figure 3

To cite this report: Monger WA, Eden RJ, 2019. First report of *Nerine latent virus* in *Haemanthus albiflos* in the UK. *New Disease Reports* **39**, 3. <http://dx.doi.org/10.5197/j.2044-0588.2019.039.003>

©2019 The Authors

This report was published on-line at www.ndrs.org.uk where high quality versions of the figures can be found.